

Genetic differences on intracranial versus spinal cord ependymal tumors: a meta-analysis of genetic researches

Chang-Hyun Lee¹ · Chun Kee Chung^{2,3,4,5} · Chi Heon Kim^{2,3,4}

Received: 25 April 2016 / Revised: 31 July 2016 / Accepted: 8 August 2016 / Published online: 16 September 2016
© Springer-Verlag Berlin Heidelberg 2016

Abstract

Purpose Although ependymomas occur in both the brain and the spine, the prognosis is quite varied by tumor location. Spinal ependymomas usually follow a relatively benign course with more favorable prognosis than that of the intracranial ependymomas. The aim of this study is to evaluate the genetic differences between spinal ependymomas and their intracranial counterparts using a meta-analysis.

Methods We searched PubMed, Embase, Web of Science, and the Cochrane library. Comparative or single arm genetic studies that enrolled patients with both intracranial and spinal ependymoma were included. The frequency of genetic aberration was calculated in each group. We calculated the odds ratio (OR) with 95 % confidence intervals (CIs) for direct comparative studies and the logit event rate (LER) and 95 % CI for single arm studies.

Results Twenty-five studies comprising of 380 spinal ependymomas and 964 intracranial ependymomas were

compared to determine the association of the genetic differences of ependymomas at different locations. There were 25 comparable genetic aberrations between spinal and intracranial ependymomas. Among the genes, the *NF2* mutation was significantly associated with the spinal ependymomas rather than with the intracranial ependymomas (spinal tumor: LER -0.750 , 95 % CI -1.233 to -0.266 , intracranial tumor: LER -3.080 , 95 % CI -3.983 to -2.177). Intracranial ependymomas were found to be significantly associated with *EPB41L3* deletion (OR 0.34; 95 % CI 0.14–0.80) and *HIC1* methylation (OR 0.12; 95 % CI 0.02–0.68).

Conclusion The genetic aberrations of spinal ependymomas are quite different from those of intracranial ependymomas. The difference in prognosis of ependymoma by location may be associated with genetic difference. A more detailed understanding of them may enable the development of targeted therapy and the estimation of prognosis.

Keywords Ependymoma · Gene · Spinal · Intracranial · Meta-analysis

✉ Chun Kee Chung
chungc@snu.ac.kr

¹ Department of Neurosurgery, Ilsan Paik Hospital, Inje University College of Medicine, Goyang, Republic of Korea

² Department of Neurosurgery, Seoul National University Hospital, Seoul National University College of Medicine, 101 Daehak-Ro, Jongno-Gu, Seoul 03080, Republic of Korea

³ Neuroscience Research Institute, Seoul National University Medical Research Center, Seoul, Republic of Korea

⁴ Clinical Research Institute, Seoul National University Hospital, Seoul, Republic of Korea

⁵ Department of Brain and Cognitive Sciences, Seoul National University College of Natural Sciences, Seoul, Republic of Korea

Introduction

Ependymomas occur in both the brain and the spine in pediatric and adult populations [1, 2]. Although ependymomas from different locations are histopathologically similar, their molecular landscape is very heterogeneous; they show differences in DNA copy number alterations, mRNA expression profiles, genetic and epigenetic alterations, and diverse transcriptional programs [1, 3–7]. The genetic landscape of ependymoma is also quite heterogeneous; these tumors mostly show complex aberration patterns with frequent deletions or gains of chromosomes,

with the deletions occurring primarily on chromosomes 1p, 4q, 6q, 9, 10, 13q, 16, 17, 19q, 20q, and 22q [8–13].

The greater difficulty in estimating the clinical course is that ependymal tumors are heterogeneous with respect to morphology, the age at which the first clinical manifestation occurs, and site-specific prognosis [14]. In children, 90 % of the ependymomas develop in the intracranial region and are associated with frequent recurrences [1, 15–17]. In adults, 60 % of the ependymomas develop in the spinal cord with rare recurrences [15, 16]. It has been previously reported that age ≥ 18 years, spinal localisation, and complete resection were positive prognosticator for the progression free survival in ependymoma [18]. These observations support the hypothesis that the histological entity “ependymoma” in fact is comprised of a group of related diseases that are likely to require different treatment approaches [4, 13, 19]. However, treatment is mainly based on surgery with or without radiation therapy, which has not been significantly changed in the last 20 years [18]. There is an urgent need for prognostic markers to tailor the treatment strategy.

The meta-analysis described here resolves the genetic differences between spinal ependymomas and their intracranial counterparts, which is essential in guiding therapeutic strategies and estimating prognosis.

Materials and methods

Literature search

We used the Embase, PubMed, Web of science, and Cochrane Library databases to undertake a comprehensive systematic literature review of all the gene studies published until March 15, 2016, with the objective to evaluate the relation of intracranial and spinal ependymomas. The following search terms were used: ependymoma, intracranial, spinal, and gene. The data were independently extracted by two reviewers who had expertise in spinal diseases and bioinformatics. In case of a discrepancy, a third author participated in the discussion until a consensus was reached. Articles in the reference lists of the selected studies were also searched manually. The search was not limited to any specific language.

Study eligibility criteria

We systematically reviewed and selected the published studies that met the following criteria: (1) case–control or cohort studies that included data regarding genetic aberrations in the patients with either intracranial or spinal intramedullary ependymoma, or both, (2) studies that provided the number of genetic aberrations and total

inspected patients, (3) studies that clearly described the DNA genotyping methods used and the sources. We excluded case reports, narrative reviews, letters, editorials, comments, biomechanical, and cadaveric studies. Studies that did not meet the eligibility criteria listed above and duplicate publications were excluded.

Data synthesis and analysis

The retrieved data included the following items: (1) study profile (author’s name, publication date, and journal), (2) study type and region, (3) participants’ demographics (such as mean age, sample size, and gender), and (4) counts of genetic aberrations in intracranial and spinal ependymomas. If the data were insufficient or unclear in a study, we attempted to contact the authors for further details.

Statistical analysis

All of the data in the direct comparative studies were dichotomous data, expressed as odds ratio (OR) of event rate with 95 % confidence interval (CI), to assess the association between genetic abnormalities and ependymomas. To analyze the relation with regard to genetic alterations, we calculated the OR with 95 % CI for comparative studies. For the single arm study, the effect size was calculated event rate as same as pooled frequency and compared with each other using the logit event rate (LER) of the standard normal (Z) distribution analysis of variance. The studies were weighted in the meta-analysis by the inverse of the variance, which included both within and between-study errors.

Between-study heterogeneity was evaluated using the inconsistency index (I^2) statistic and Chi square (χ^2), with the statistical significance set at $P < 0.10$. When the heterogeneity was not statistically significant and the value of I^2 was less than 50 %, the fixed-effects model was used to estimate the pooled OR or LER. The funnel plot and Egger’s test were performed to assess publication bias. The overall effect was tested by Fisher’s z transformation (statistical significance set at $P < 0.05$). Sensitivity analysis was performed by varying assumptions used in the meta-analysis and by single elimination of the studies. If there was no mutation in all groups (i.e., a “zero cell” in the 2 by 2 table), 0.5 was added to each cell so that the estimated values would not be 0 or infinity and so that the standard error could be calculated. Statistical analyses were performed using Review Manager (RevMan) Version 5.3. (The Nordic Cochrane Centre, the Cochrane Collaboration, Copenhagen, Denmark), comprehensive meta-analysis, version 3.0 (Biostat, Englewood, NJ, USA), and R software (version 3.2.0, The R Foundation for Statistical Computing, Vienna, Austria).

Results

Based on the selection strategy described in the methods section, a total of 79 articles were identified (PubMed, 21; EMBASE, 34; Web of Science, 24; Cochrane library, 0). After screening and reviewing for eligibility, a total of 25 studies, including 380 spinal and 964 intracranial ependymomas, were retrieved for this meta-analysis. The detailed selection process is shown in Fig. 1. We sent seven corresponding authors e-mail to request their data twice, none answered the request.

The baseline characteristics of the studies included in this meta-analysis are presented in Table 1. Of the 25 articles we identified, the study/studies conducted in the USA [15, 20–25], Europe [6, 14, 26–39], and Asia [40, 41] were 7, 16, and 2, respectively. In all these studies, the patients with intracranial ependymoma were much younger than those with spinal ependymoma in Table 1. Intracranial ependymomas usually showed male predominance, consistent with a previous epidemiologic report [42]; however, spinal ependymomas did not demonstrated such clear sex-predominance.

Six papers investigated the association of *NF2* gene with ependymoma susceptibility. The pooled frequency of *NF2* mutations in spinal and intracranial ependymoma was 32.1 and 4.4 % in Table 2, respectively. Comparison performing using the LER of forest plots were significantly different in mutation frequency between the two groups as shown in Fig. 2. According to χ^2 statistic (intracranial, $P = 0.63$; spinal, $P = 0.21$) and I^2 statistic (intracranial, $I^2 = 0\%$; spinal, $I^2 = 31.36\%$), heterogeneity was assessed and found not to be significant, therefore, a fixed-effect approach was used to determine pooled frequency of *NF2* mutation of spinal versus intracranial ependymomas.

EPB41L3 deletion accounted for 46.6 % of intracranial ependymoma and 21.7 % of spinal ependymoma (OR 0.34; 95 % CI 0.14–0.80; $P = 0.01$) and the *EPB41L3* deletion is substantially associated with intracranial ependymoma compared with spinal ependymoma in Fig. 3. The pooled frequency of *HIC1* hypermethylation was 93.8 % of intracranial ependymoma and 65.0 % of spinal ependymoma, which displayed the close association between intracranial ependymoma and *HIC1* hypermethylation (OR 0.12; 95 % CI 0.02–0.68; $P = 0.02$).

The other gene mutations in the following genes did not show a predominant association with tumors at any location: *NEFL*, *CDKN2A*, *EGFR*, *HOXB13*, *PDGFRA*, *RASSF1*, *RB*, *SMARCB1*, *CDKN2B*, *TNC*, *CASP8*, *MOS*, *MEN1*, *NOTCH1*, *SHC3* and *SIPR3*, *THBS1*, *PTEN*, *TP73*, *MGMT*, *TIMP3*, *ERBB2*, and *MDM2* genes as shown in Table 2 and Fig. 3. The pooled frequency of *NEFL* overexpression in spinal and intracranial ependymoma was 59.1 % (95 % CI 42.9–73.6 %) and 38.3 % (95 % CI 27.8–50.0 %) in Table 2, respectively. The gene did not show a substantial association with the spinal ependymomas in Fig. 4. Among 22 myxopapillary ependymomas in the spinal cord tumor, *NEFL* overexpression accounted for 69.0 % (95 % CI 44.1–86.3 %) of the tumor, which demonstrated no substantial difference in frequency of *NEFL* mutation between myxopapillary ependymoma and the other ependymomas.

Ten studies investigated *CDKN2A* mutations in intracranial and/or spinal ependymoma. *CDKN2A* mutations were observed in 20.6 % in intracranial ependymoma and 21.9 % of spinal ependymoma in Table 2. LER of intracranial and spinal ependymoma was -1.347 and -1.121 , respectively, in Fig. 5. The pooled frequency of

Fig. 1 A flow chart showing the process of study identification and exclusion

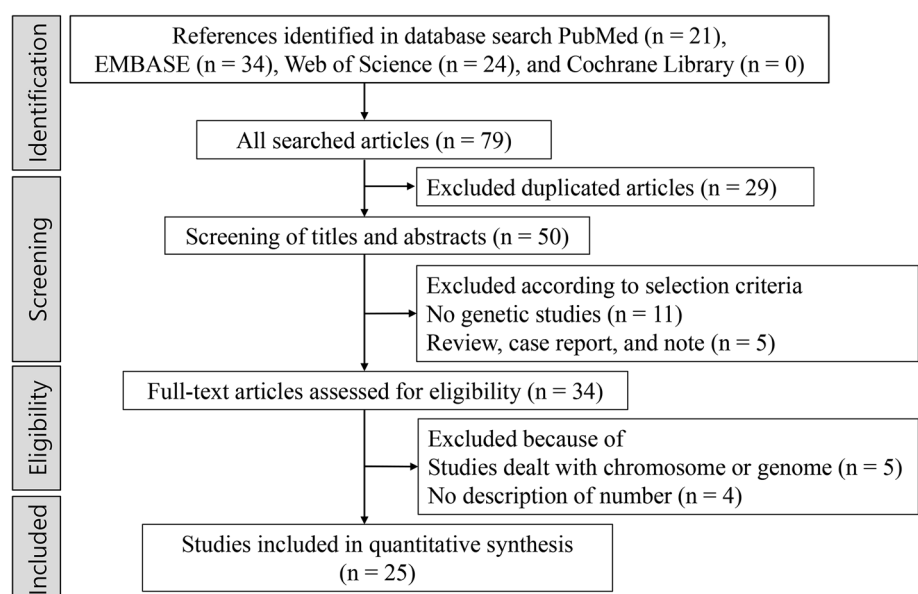


Table 1 The baseline characteristics of the included studies

References	Study design	Study region	Number		Mean age		Male (%)	
			IC	Sp	IC	Sp	IC	Sp
Ebert et al. [14]	Retro	Germany	39	23	24	41	51	65
Suzuki et al. [41]	Retro	Japan	16	4	23	37	75	100
Lamszus et al. [33]	Retro	Germany	20	32	21	39	86	58
Bortolotto et al. [29]	Retro	Italy	16	0	N/D	N/D	N/D	N/D
Kraus et al. [32]	Retro	Germany	31	17	26	40	N/D	N/D
Singh et al. [15]	Retro	USA	12	15	17	39	86	29
Gilberston et al. [30]	Retro	UK	120	1	N/D	N/D	N/D	N/D
Rousseau et al. [38]	Retro	Belgium	60	43	N/D	N/D	N/D	N/D
Athanasίου et al. [28]	Retro	UK	29	5	N/D	N/D	N/D	N/D
Rajaram et al. [20]	Retro	USA	50	51	16	41	N/D	N/D
Alonso et al. [26]	Retro	Spain	2	25	18	31	56	50
Waha et al. [39]	Retro	Germany	35	20	16	39	38	48
Hamilton et al. [31]	Retro	UK	9	11	N/D	N/D	N/D	N/D
Rajaram et al. [25]	Retro	USA	51	33	7 ^a	41 ^a	N/D	N/D
Mendrzyk et al. [35]	Retro	Germany	49	19	N/D	N/D	N/D	N/D
Michalowski et al. [36]	Retro	France	27	0	N/D	N/D	N/D	N/D
Andreiuolo et al. [18]	Retro	France	66	0	N/D	N/D	45	N/D
Barton et al. [22]	Retro	USA	23	5	3 ^a	13 ^a	65	25
Magrassi et al. [34]	Retro	Italy	16	19	15	44	69	47
Korshunov et al. [6]	Retro	Germany	122	0	N/D	N/D	48	N/D
Modena et al. [37]	Retro	Italy	60	0	3	N/D	57	N/D
Stephen et al. [23]	Retro	USA	0	19	N/D	13	N/D	63
Bettegowda et al. [21]	Retro	USA	40	19	N/D	N/D	N/D	N/D
Karakoula et al. [24]	Retro	USA	40	0	N/D	N/D	N/D	N/D
Gupta et al. [40]	Retro	India	31	19	20	32	71	68

IC intracranial ependymoma, Sp spinal ependymoma, Retro retrospective study, N/D not described

^a Median age

Table 2 Pooled frequency (event rate) of genetic mutations in ependymoma of single group studies

Locus	Gene	Intracranial ependymoma		Spinal ependymoma	
		Event rate (%)	95 % CI (%)	Event rate (%)	95 % CI (%)
22q12	NF2 mutation	4.4	1.8–10.2	32.1	22.6–43.4
8p21	NEFL overexpression	38.3	27.8–50.0	59.1	42.9–73.6
9p21	CDKN2A deletion	20.6	16.4–25.6	21.9	15.9–29.4
7p11	EGFR overexpression	52.1	38.5–65.3	29.8	13.0–54.5
17q21	HOXB13 overexpression	22.2	5.6–57.9	59.0	39.5–76.0
4q12	PDGFRA overexpression	84.6	54.9–96.1	83.8	68.3–92.5
3p21	RASSF1 methylation	64.5	47.7–78.4	90.0	67.6–97.5
13q14	RB deletion	15.9	9.2–26.2	23.3	14.0–36.2
22q11	SMARCB1 deletion	15.3	6.7–31.2	21.4	7.1–49.4

Event rate showed significant difference between two groups

CDKN2A mutations was slightly higher in the spinal ependymoma than that in the intracranial counterpart, which is not substantially different.

TNC mutation was frequently observed in infratentorial ependymoma, in comparison to spinal and supratentorial

ependymomas. The expression frequency of *TNC* mutation in infratentorial ependymoma was reported to be 50.0 %, whereas it was 18.8 and 31.2 % in spinal and supratentorial ependymomas, respectively [40]. *NOTCH1* mutation in supratentorial tumors was significantly higher (73 %)

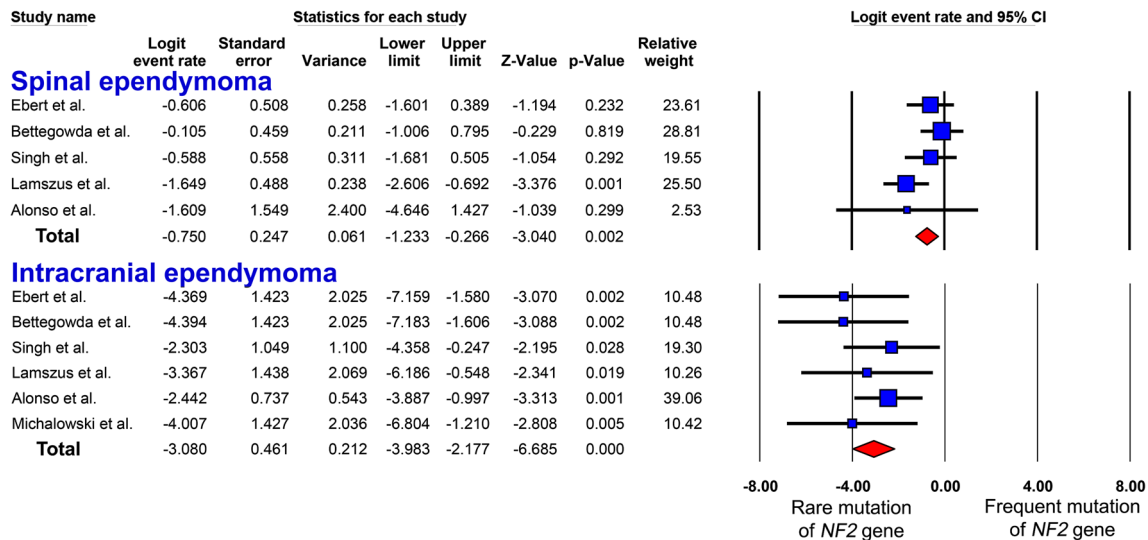


Fig. 2 Forest plot of the logit event rate of *NF2* mutation by study and subgroup in the spinal and intracranial ependymoma group. Spinal ependymoma shows substantially frequent mutation of *NF2* gene compared with the intracranial ependymoma

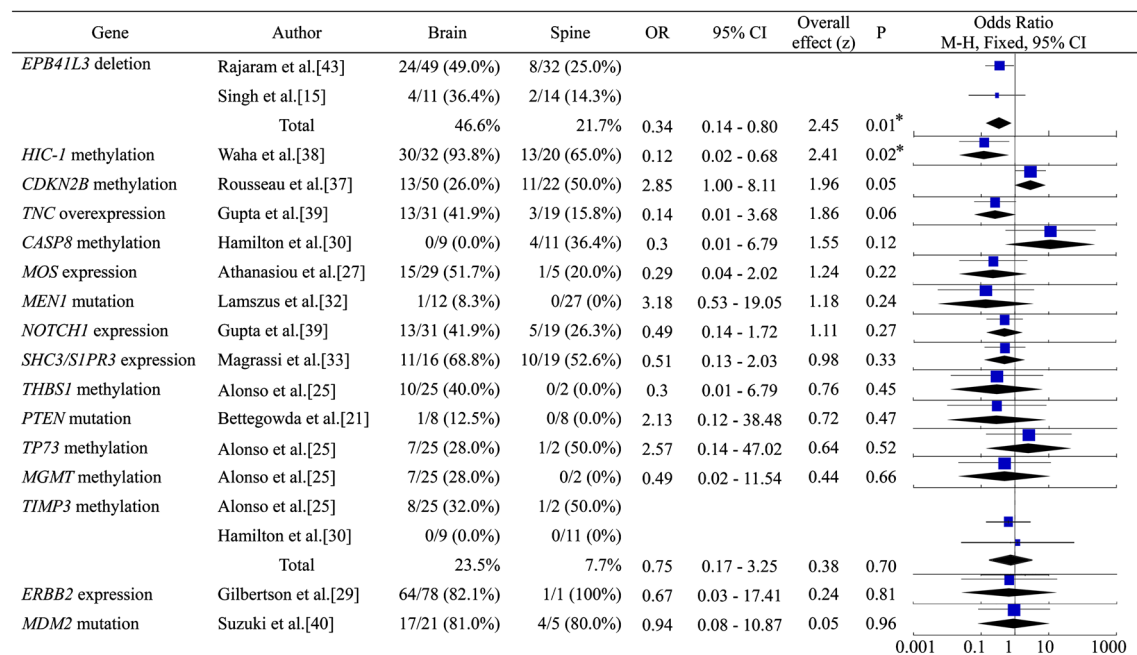


Fig. 3 Pooled frequency of genetic mutations in ependymoma of direct comparative studies. *NEPB41L3* and *HIC1* aberrations show substantially different association between intracranial and spinal

ependymoma. *Odds ratio shows significant difference in the pooled frequency of mutation between two groups

than in infratentorial (19 %; $P = 0.001$) and spinal (26 %; $P = 0.01$) tumors [40]. *MEN1* and *PTEN* mutations have been rarely observed in ependymomas [14, 21, 33]. Comparisons of more than three studies demonstrated usually minimal to moderate heterogeneity in Table 3. Funnel plots and the Egger test showed a little risk of publication bias except intracranial *RB* mutation. It calculated as -2.27 ($P = 0.04$), which may indicate an underpowered analysis.

Discussion

Although pathological findings of spinal and intracranial ependymomas are similar to each other, their clinical course and commonly affected age are quite different. Genetic difference can be a possible reason of this discrepancy and this meta-analysis displayed that spinal ependymoma is associated with *NF2* whereas intracranial ependymoma is associated with *EPB41L3* and *HIC1*. The

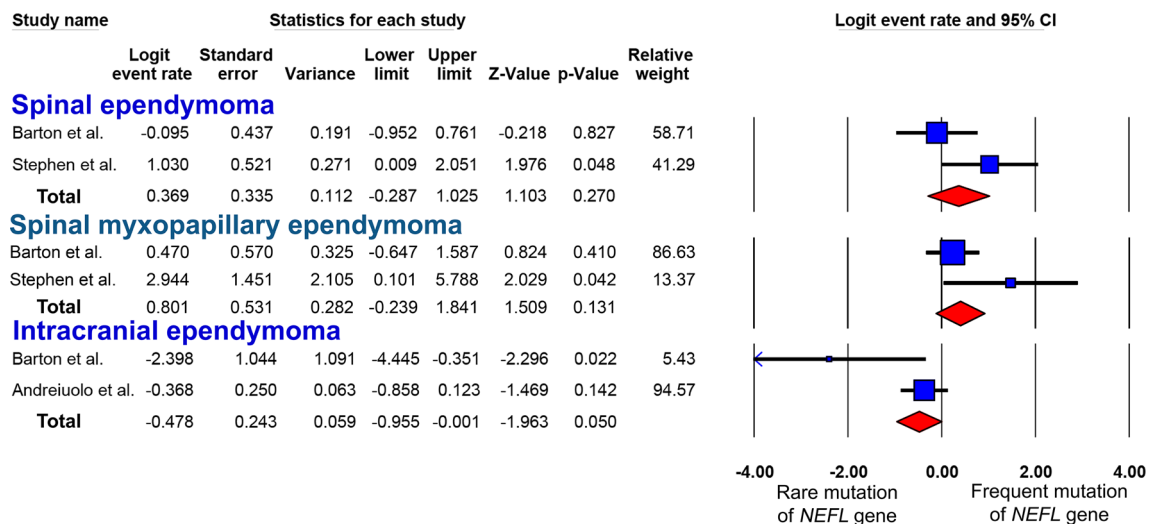


Fig. 4 Forest plot of the logit event rate of *NEFL* mutation by study and subgroup in the spinal and intracranial ependymoma group. *NEFL* mutation is frequent in myxopapillary ependymoma. However, there is no significant difference in frequency of mutation. *CI* confidence interval

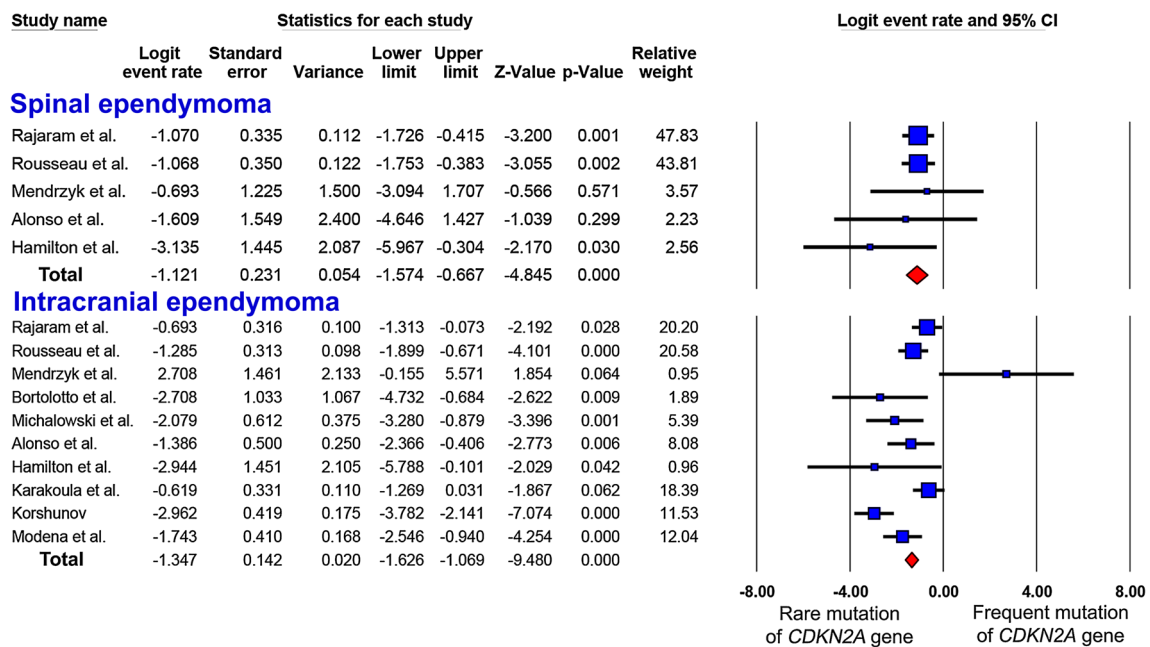


Fig. 5 Forest plot of the logit event rate of *CDKN2A* mutation by study and subgroup in the spinal and intracranial ependymoma group. Both intracranial and spinal ependymoma shows low to moderate frequency of *CDKN2A* mutation. *CI* confidence interval

Table 3 Statistical tests for heterogeneity and publication bias

Gene	Location	χ^2 (P value)	I^2	τ^2	Egger (P value)
<i>NF2</i> mutation	Intracranial E	3.43 (0.63)	0.0	0.0	-2.36 (0.23)
	Spinal E	5.83 (0.21)	31.36	0.15	-0.90 (0.68)
<i>CDKN2A</i> deletion	Intracranial E	31.68 (0.00)	74.74	0.73	-0.31 (0.85)
	Spinal E	2.17 (0.54)	0.0	0.0	-0.69 (0.44)
<i>RB</i> deletion	Intracranial E	7.00 (0.07)	57.09	1.20	-2.27 (0.04)
	Spinal E	2.02 (0.37)	0.72	0.01	-1.13 (0.37)

other 22 genes did not show substantial difference in frequency of mutation by tumor location.

Although a variety of numerical and structural chromosomal abnormalities have been shown to be associated with ependymomas, inactivation of the *NF2* gene as well as the sporadic mutations in *NF2* have been well documented in spinal ependymomas [7, 14, 15, 43]. The importance of the *NF2* gene in spinal ependymoma pathogenesis is further emphasized by the observation that *NF2* mutations and the loss of expression of Merlin (the protein encoded by the *NF2* gene) are found in 30–71 % of sporadic (non-syndromic) ependymomas [15, 44]. Among six enrolled studies on *NF2* included in our meta-analysis, four studies reported that *NF2* mutation of intracranial ependymoma was not observed at all among 120 patients [14, 21, 33, 36]. Another study reported that *NF2* loss was only a trend at the DNA level, whereas it was highly clearly associated with spinal ependymomas at the protein (Merlin) level [15, 45].

Although *NF2* loss was frequently observed in spinal ependymoma, pooled frequency of *NF2* mutation in spinal ependymoma was 32.1 %. The low frequency of genetic mutation may be explained by dilution of high and low frequency. A previous studies addressed that *NF2* mutations were observed in a high percentage of spinal ependymoma (WHO grade II), but in low percentage of intracranial WHO grade II ependymomas as well as all WHO grade I myxopapillary ependymoma, WHO grade I subependymoma, and WHO grade III ependymoma studied [14]. Because of insufficient data, this study could not perform subgroup analysis by WHO grade. Further clinical study is needed to determine the relation between specific genetic mutation and ependymoma.

This analysis revealed that intracranial ependymomas clearly demonstrated *EPB41L3* and *HIC1* mutations, in comparison to their spinal counterparts [15, 25, 39]. Two studies included a total of 106 patients investigated *EPB41L3* genes in ependymomas and one study of 52 patients dealt with *HIC1* mutation. Although the difference in frequency of these genes mutation was statistically significant, further evaluation and large study are needed because only a few studies investigated *HIC1* and *EPB41L3* genes.

Myxopapillary ependymomas may be considered molecularly different from intracranial ependymomas as well as other spinal ependymomas [22]. Previous studies reported that they showed high expression of some genes, including *NEFL*, *HOXB5*, *PLA2G5*, and *ITIH2* [22]. We performed subgroup analysis of myxopapillary ependymoma for all reported genes. However, no unique genetic mutation of myxopapillary ependymoma was observed. Among evaluated genes, we showed the event rate of *NEFL* aberration because *NEFL* is known to the common genetic mutation of myxopapillary ependymoma. This

meta-analysis showed that *NEFL* overexpression was 69.0 % in spinal myxopapillary ependymomas, 53.8 % in other spinal ependymomas (except myxopapillary ependymoma), and 38.3 % in intracranial ependymoma. Myxopapillary ependymoma showed a tendency of high incidence of *NEFL* mutation, but it did not show statistical differences in frequency.

Some review papers dealt with genetics of ependymomas [45–47]. The review papers simply described possible genes, proteins, and chromosomal aberrations in spinal, intracranial ependymoma, or both. They reported the differences between spinal and intracranial ependymoma in aspect of gene, protein, and chromosome. They addressed the frequency of genetic aberrations of each comparative study between brain and spine region. In contrast, this meta-analysis included both the comparative studies and single region studies in brain or spinal cord lesion, and summarized pooled effect size of genetic mutation. It may provide more precise perspective in aspect of gene than the review papers.

Currently, the only effective treatment for clinically symptomatic ependymoma is surgery, with or without radiotherapy [44]. Such surgical procedures, however, pose substantial risks, such as worsening of neurological deficits, paralysis, and death [16]. Despite active genetic researches, targeted therapeutics for ependymomas is in very early stage of its development. The currently used targeted therapeutic approaches depend on the similarities of ependymomas with other glial tumors, rather than influencing pathways specific to ependymomas. Merlin regulates the growth of the spinal cord neural progenitor cells in a Rac1- and ErbB2-dependent manner, which firmly establishes a central growth control target for *NF2*-associated spinal ependymoma [44, 48]. *NF2*-associated spinal ependymomas exhibit increased ErbB2 activation, resulting from Rac1-mediated ErbB2 retention on the plasma membrane [44]. Lapatinib, a selective ErbB1 and ErbB2 inhibitor prolonged disease stabilization in patients with ependymoma, in the phase I study [49]. Based on these encouraging data, a phase II study of bevacizumab and lapatinib in children with recurrent or refractory ependymomas was conducted [50]. Unfortunately, the combination therapy proved ineffective in treating recurrent ependymomas. Although this meta-analysis cannot demonstrate the reliable treatment of ependymoma based on the genetic perspective, it may help understanding ependymoma and be a cornerstone of ependymoma treatment.

Limitations of our study

Our study has some limitations that must be considered when interpreting our results. Firstly, there are potential pitfalls which could distort the results in weakness and

biases of genetic studies, such as study design, genotyping error, and population stratification [51]. This bias of genetic studies is related with enrolled primary studies. It is regarded to be corrected by replication studies. The bias of this meta-analysis may be low because the meta-analysis enrolled all relative studies and produce the summary of estimate. Each study verified genetic aberrations in gene or protein level using different tools, such as comparative genomic hybridization (CGH), fluorescence in situ hybridization (FISH), and immunohistochemistry (IHC). Heterogeneity in the design and clinical characteristics of the studied populations might also have contributed to slightly attenuate the power of our study to detect additional significant associations. Secondly, publication bias is another potentially important limitation of this study. Although neither Egger's test nor Begg's test found evidence for publication bias in this meta-analysis, negative studies are less likely to be published indeed, which may affect the validity of analysis [52]. Besides, few papers dealt with *EPB41L3* or *HIC1* genes. So, considering the potential limitations of studies included in current meta-analysis, our results should be interpreted with caution.

Conclusion

The genetic landscape of spinal ependymomas is quite different from that of intracranial ependymomas; the former seems to be closely associated with *NF2* mutation, whereas the latter seems to be associated with *EPB41L3* deletion and *HIC1* methylation. A more detailed understanding of these various genetic aberrations may enable the identification of more specific prognostic markers, as well as the development of customized and targeted therapy.

Acknowledgments This work was supported by the National Research Foundation of Korea (NRF) Grant funded by the Korea government (MSIP) (No. 2010-0028631). The sponsor had no role in the design or conduct of this research.

Compliance with ethical standards

Conflicts of interest None of the authors has any financial interest in the subject under discussion in this paper.

References

- Wani K, Armstrong TS, Vera-Bolanos E, Raghunathan A, Ellison D, Gilbertson R, Vaillant B, Goldman S, Packer RJ, Fouladi M, Pollack I, Mikkelsen T, Prados M, Omuro A, Soffiatti R, Ledoux A, Wilson C, Long LH, Gilbert MR, Aldape K, Collaborative Ependymoma R (2012) A prognostic gene expression signature in infratentorial ependymoma. *Acta Neuropathol* 123:727–738
- Vera-Bolanos E, Aldape K, Yuan Y, Wu J, Wani K, Necesito-Reyes MJ, Colman H, Dhall G, Lieberman FS, Metellus P, Mikkelsen T, Omuro A, Partap S, Prados M, Robins HI, Soffiatti R, Wu J, Gilbert MR, Armstrong TS, Foundation C (2015) Clinical course and progression-free survival of adult intracranial and spinal ependymoma patients. *Neuro Oncol* 17:440–447
- Modena P, Lualdi E, Facchinetti F, Veltman J, Reid JF, Minardi S, Janssen I, Giangaspero F, Forni M, Finocchiaro G, Genitori L, Giordano F, Riccardi R, Schoenmakers E, Massimino M, Sozzi G (2006) Identification of tumor-specific molecular signatures in intracranial ependymoma and association with clinical characteristics. *J Clin Oncol* 24:5223–5233
- Taylor MD, Poppleton H, Fuller C, Su XP, Liu YX, Jensen P, Magdaleno S, Dalton J, Calabrese C, Board J, MacDonald T, Rutka J, Guha A, Gajjar A, Curran T, Gilbertson RJ (2005) Radial glia cells are candidate stem cells of ependymoma. *Cancer Cell* 8:323–335
- Puget S, Grill J, Valent A, Bieche I, Dantas-Barbosa C, Kauffmann A, Dessen P, Lacroix L, Georger B, Job B, Dirven C, Varlet P, Peyre M, Dirks PB, Sainte-Rose C, Vassal G (2009) Candidate genes on chromosome 9q33–34 involved in the progression of childhood ependymomas. *J Clin Oncol* 27:1884–1892
- Korshunov A, Witt H, Hielscher T, Benner A, Remke M, Ryzhova M, Milde T, Bender S, Wittmann A, Schottler A, Kulozik AE, Witt O, von Deimling A, Lichter P, Pfister S (2010) Molecular staging of intracranial ependymoma in children and adults. *J Clin Oncol* 28:3182–3190
- Pajtler KW, Witt H, Sill M, Jones DTW, Hovestadt V, Kratochwil F, Wani K, Tatevossian R, Punchihewa C, Johann P, Reimand J, Warnatz HJ, Ryzhova M, Mack S, Ramaswamy V, Capper D, Schweizer L, Sieber L, Wittmann A, Huang Z, van Sluis P, Volckmann R, Koster J, Versteeg R, Fults D, Toledano H, Avigad S, Hoffman LM, Donson AM, Foreman N, Hower E, Zitterbart K, Gilbert M, Armstrong TS, Gupta N, Allen JC, Karajannis MA, Zagzag D, Hasselblatt M, Kulozik AE, Witt O, Collins VP, von Hoff K, Rutkowski S, Pietsch T, Bader G, Yaspo ML, von Deimling A, Lichter P, Taylor MD, Gilbertson R, Ellison DW, Aldape K, Korshunov A, Kool M, Pfister SM (2015) Molecular classification of ependymal tumors across all CNS compartments, histopathological grades, and age groups. *Cancer Cell* 27:728–743
- Huang B, Starostik P, Schraut H, Krauss J, Sorensen N, Roggendorf W (2003) Human ependymomas reveal frequent deletions on chromosomes 6 and 9. *Acta Neuropathol* 106:357–362
- Scheil S, Bruderlein S, Eicker M, Herms J, Herold-Mende C, Steiner HH, Barth TFE, Moller P (2001) Low frequency of chromosomal imbalances in anaplastic ependymomas as detected by comparative genomic hybridization. *Brain Pathol* 11:133–143
- Ward S, Harding B, Wilkins P, Harkness W, Hayward R, Darling JL, Thomas DGT, Warr T (2001) Gain of 1q and loss of 22 are the most common changes detected by comparative genomic hybridisation in paediatric ependymoma. *Genes Chromosomes Cancer* 32:59–66
- Zheng PP, Pang JC, Hui AB, Ng HK (2000) Comparative genomic hybridization detects losses of chromosomes 22 and 16 as the most common recurrent genetic alterations in primary ependymomas. *Cancer Genet Cytogenet* 122:18–25
- Witt H, Mack SC, Ryzhova M, Bender S, Sill M, Isserlin R, Benner A, Hielscher T, Milde T, Remke M, Jones DT, Northcott PA, Garzia L, Bertrand KC, Wittmann A, Yao Y, Roberts SS, Massimi L, Van Meter T, Weiss WA, Gupta N, Grajkowska W, Lach B, Cho YJ, von Deimling A, Kulozik AE, Witt O, Bader GD, Hawkins CE, Tabori U, Guha A, Rutka JT, Lichter P, Korshunov A, Taylor MD, Pfister SM (2011) Delineation of two

- clinically and molecularly distinct subgroups of posterior fossa ependymoma. *Cancer Cell* 20:143–157
13. Johnson RA, Wright KD, Poppleton H, Mohankumar KM, Finkelstein D, Pounds SB, Rand V, Leary SES, White E, Eden C, Hogg T, Northcott P, Mack S, Neale G, Wang YD, Coyle B, Atkinson J, DeWire M, Kranenburg TA, Gillespie Y, Allen JC, Merchant T, Boop FA, Sanford RA, Gajjar A, Ellison DW, Taylor MD, Grundy RG, Gilbertson RJ (2010) Cross-species genomics matches driver mutations and cell compartments to model ependymoma. *Nature* 466:632–636
 14. Ebert C, von Haken M, Meyer-Puttitz B, Wiestler OD, Reifenberger G, Pietsch T, von Deimling A (1999) Molecular genetic analysis of ependymal tumors. NF2 mutations and chromosome 22q loss occur preferentially in intramedullary spinal ependymomas. *Am J Pathol* 155:627–632
 15. Singh PK, Gutmann DH, Fuller CE, Newsham IF, Perry A (2002) Differential involvement of protein 4.1 family members DAL-1 and NF2 in intracranial and intraspinal ependymomas. *Mod Pathol* 15:526–531
 16. Zadnik PL, Gokaslan ZL, Burger PC, Bettegowda C (2013) Spinal cord tumours: advances in genetics and their implications for treatment. *Nat Rev Neurol* 9:257–266
 17. Kilday JP, Rahman R, Dyer S, Ridley L, Lowe J, Coyle B, Grundy R (2009) Pediatric ependymoma: biological perspectives. *Mol Cancer Res* 7:765–786
 18. Andreiulo F, Ferreira C, Puget S, Grill J (2013) Current and evolving knowledge of prognostic factors for pediatric ependymomas. *Future Oncol* 9:183–191
 19. Parker M, Mohankumar KM, Punchihewa C, Weinlich R, Dalton JD, Li YJ, Lee R, Tatevossian RG, Phoenix TN, Thiruvengadam R, White E, Tang B, Orisme W, Gupta K, Rusch M, Chen X, Li YX, Nagahawhatte P, Hedlund E, Finkelstein D, Wu G, Shurtleff S, Easton J, Boggs K, Yergeau D, Vadodaria B, Mulder HL, Becksford J, Gupta P, Huether R, Ma J, Song GC, Gajjar A, Merchant T, Boop F, Smith AA, Ding L, Lu C, Ochoa K, Zhao D, Fulton RS, Fulton LL, Mardis ER, Wilson RK, Downing JR, Green DR, Zhang JH, Ellison DW, Gilbertson RJ (2014) C11orf95-RELA fusions drive oncogenic NF-kappa B signalling in ependymoma. *Nature* 506:451–455
 20. Rajaram V, Leuthardt EC, Singh PK, Ojemann JG, Brat DJ, Prayson RA, Perry A (2003) 9p21 and 13q14 dosages in ependymomas. A clinicopathologic study of 101 cases. *Mod Pathol* 17:9–14
 21. Bettegowda C, Agrawal N, Jiao YC, Wang YX, Wood LD, Rodriguez FJ, Hruban RH, Gallia GL, Binder ZA, Riggins CJ, Salmasi V, Riggins GJ, Reitman ZJ, Rasheed A, Keir S, Shinjo S, Marie S, McLendon R, Jallo G, Vogelstein B, Bigner D, Yan H, Kinzler KW, Papadopoulos N (2013) Exomic sequencing of four rare central nervous system tumor types. *Oncotarget* 4:572–583
 22. Barton VN, Donson AM, Kleinschmidt-DeMasters BK, Birks DK, Handler MH, Foreman NK (2010) Unique molecular characteristics of pediatric myxopapillary ependymoma. *Brain Pathol* 20:560–570
 23. Stephen JH, Sievert AJ, Madsen PJ, Judkins AR, Resnick AC, Storm PB, Rushing EJ, Santi M (2012) Spinal cord ependymomas and myxopapillary ependymomas in the first 2 decades of life: a clinicopathological and immunohistochemical characterization of 19 cases. *J Neurosurg Pediatrics* 9:646–653
 24. Karakoula K, Jacques TS, Phipps KP, Harkness W, Thompson D, Harding BN, Darling JL, Warr TJ (2014) Epigenetic genome-wide analysis identifies BEX1 as a candidate tumour suppressor gene in paediatric intracranial ependymoma. *Cancer Lett* 346:34–44
 25. Rajaram V, Gutmann DH, Prasad SK, Mansur DB, Perry A (2005) Alterations of protein 4.1 family members in ependymomas: a study of 84 cases. *Mod Pathol* 18:991–997
 26. Alonso ME, Bello MJ, Gonzalez-Gomez P, Arjona D, de Campos JM, Gutierrez M, Rey JA (2004) Aberrant CpG island methylation of multiple genes in ependymal tumors. *J Neurooncol* 67:159–165
 27. Andreiulo F, Puget S, Peyre M, Dantas-Barbosa C, Boddaert N, Philippe C, Mauguén A, Grill J, Varlet P (2010) Neuronal differentiation distinguishes supratentorial and infratentorial childhood ependymomas. *Neuro Oncol* 12:1126–1134
 28. Athanasiou A, Perunovic B, Quilty RD, Gorgoulis VG, Kittas C, Love S (2003) Expression of *mos* in ependymal gliomas. *Am J Clin Pathol* 120:699–705
 29. Bortolotto S, Chiado-Piat L, Cavalla P, Bosone I, Mauro A, Schiffer D (2001) CDKN2A/p16 in ependymomas. *J Neurooncol* 54:9–13
 30. Gilbertson RJ, Bentley L, Hernan R, Junttila TT, Frank AJ, Haapasalo H, Connelly M, Wetmore C, Curran T, Elenius K, Ellison DW (2002) ERBB receptor signaling promotes ependymoma cell proliferation and represents a potential novel therapeutic target for this disease. *Clin Cancer Res* 8:3054–3064
 31. Hamilton DW, Lusher ME, Lindsey JC, Ellison DW, Clifford SC (2005) Epigenetic inactivation of the RASSF1A tumour suppressor gene in ependymoma. *Cancer Lett* 227:75–81
 32. Kraus JA, de Millas W, Sorensen N, Herbold C, Schichor C, Tonn JC, Wiestler OD, von Deimling A, Pietsch T (2001) Indications for a tumor suppressor gene at 22q11 involved in the pathogenesis of ependymal tumors and distinct from hSNF5/INI1. *Acta Neuropathol* 102:69–74
 33. Lamszus K, Lachenmayer L, Heinemann U, Kluwe L, Finckh U, Hoppner W, Stavrou D, Fillbrandt R, Westphal M (2001) Molecular genetic alterations on chromosomes 11 and 22 in ependymomas. *Int J Cancer* 91:803–808
 34. Magrassi L, Marziliano N, Inzani F, Cassini P, Chiaranda I, Skrap M, Pizzolito S, Arienta C, Arbustini E (2010) EDG3 and SHC3 on chromosome 9q22 are co-amplified in human ependymomas. *Cancer Lett* 290:36–42
 35. Mendrzyk F, Korshunov A, Benner A, Toedt G, Pfister S, Radlwimmer B, Lichter P (2006) Identification of gains on 1q and epidermal growth factor receptor overexpression as independent prognostic markers in intracranial ependymoma. *Clin Cancer Res* 12:2070–2079
 36. Michalowski MB, de Fraipont F, Michelland S, Entz-Werle N, Grill J, Pasquier B, Favrot MC, Plantaz D (2006) Methylation of RASSF1A and TRAIL pathway-related genes is frequent in childhood intracranial ependymomas and benign choroid plexus papilloma. *Cancer Genet Cytogenet* 166:74–81
 37. Modena P, Buttarelli FR, Miceli R, Piccinin E, Baldi C, Antonelli M, Morra I, Lauriola L, Di Rocco C, Garre ML, Sardi I, Genitori L, Maestro R, Gandola L, Facchinetti F, Collini P, Sozzi G, Giangaspero F, Massimino M (2012) Predictors of outcome in an AIEOP series of childhood ependymomas: a multifactorial analysis. *Neuro Oncol* 14:1346–1356
 38. Rousseau E, Ruchoux MM, Scaravilli F, Chapon F, Vinchon M, De Smet C, Godfraind C, Viskula M (2003) CDKN2A, CDKN2B and p14(ARF) are frequently and differentially methylated in ependymal tumours. *Neuropathol Appl Neurobiol* 29:574–583
 39. Waha A, Koch A, Hartmann W, Mack H, Schramm J, Sorensen N, Berthold F, Wiestler OD, Pietsch T, Waha A (2004) Analysis of HIC-1 methylation and transcription in human ependymomas. *Int J Cancer* 110:542–549
 40. Gupta RK, Sharma MC, Suri V, Kakkar A, Singh M, Sarkar C (2014) Study of chromosome 9q gain, Notch pathway regulators and Tenascin-C in ependymomas. *J Neurooncol* 116:267–274
 41. Suzuki SO, Iwaki T (2000) Amplification and overexpression of *mdm2* gene in ependymomas. *Mod Pathol* 13:548–553
 42. Ostrom QT, Gittleman H, Fulop J, Liu M, Blanda R, Kromer C, Wolinsky Y, Kruchko C, Barnholtz-Sloan JS (2015) CBTRUS

- statistical report: primary brain and central nervous system tumors diagnosed in the United States in 2008–2012. *Neuro Oncol* 17 Suppl 4:iv1–iv62
43. Rubio MP, Correa KM, Ramesh V, MacCollin MM, Jacoby LB, Von Deimling A, Gusella JF, Louis DN (1994) Analysis of the neurofibromatosis 2 gene in human ependymomas and astrocytomas. *Cancer Res* 54:45–47
 44. Garcia C, Gutmann DH (2014) Nf2/Merlin controls spinal cord neural progenitor function in a Rac1/ErbB2-dependent manner. *PLoS One* 9:e97320
 45. Lee CH, Chung CK, Ohn JH, Kim CH (2016) The similarities and differences between intracranial and spinal ependymomas: a review from a genetic research perspective. *J Korean Neurosurg Soc* 59:83–90
 46. Yang I, Nagasawa DT, Kim W, Spasic M, Trang A, Lu DC, Martin NA (2012) Chromosomal anomalies and prognostic markers for intracranial and spinal ependymomas. *J Clin Neurosci* 19:779–785
 47. Yao Y, Mack SC, Taylor MD (2011) Molecular genetics of ependymoma. *Chin J Cancer* 30:669–681
 48. Hanemann CO (2008) Magic but treatable? Tumours due to loss of merlin. *Brain* 131:606–615
 49. Fouladi M, Stewart CF, Blaney SM, Onar-Thomas A, Schaiquevich P, Packer RJ, Gajjar A, Kun LE, Boyett JM, Gilbertson RJ (2010) Phase I trial of lapatinib in children with refractory CNS malignancies: a Pediatric Brain Tumor Consortium study. *J Clin Oncol* 28:4221–4227
 50. DeWire M, Fouladi M, Turner DC, Wetmore C, Hawkins C, Jacobs C, Yuan Y, Liu D, Goldman S, Fisher P, Rytting M, Bouffet E, Khakoo Y, Hwang EI, Foreman N, Stewart CF, Gilbert MR, Gilbertson R, Gajjar A (2015) An open-label, two-stage, phase II study of bevacizumab and lapatinib in children with recurrent or refractory ependymoma: a collaborative ependymoma research network study (CERN). *J Neurooncol* 123:85–91
 51. Daly AK, Day CP (2001) Candidate gene case-control association studies: advantages and potential pitfalls. *Br J Clin Pharmacol* 52:489–499
 52. Thornton A, Lee P (2000) Publication bias in meta-analysis: its causes and consequences. *J Clin Epidemiol* 53:207–216